

# Control of Deformed Wing Virus–B in a Commercial Apiary Following Queen Vaccination with an Experimental *Paenibacillus larvae* Bacterin

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Deformed Wing Virus (DWV) is a viral disease present in the majority of honey bee colonies worldwide. In addition to vertical transmission, DWV is vectored by *Varroa destructor* mites, which facilitate virulence through suppression of the bees' immune system. DWV and *Varroa* are both strongly linked to colony losses and are of great concern to beekeepers. There is no available treatment or preventative for DWV, therefore management of the disease is limited to indirect mite control. In a 400-hive placebo-controlled field study, we demonstrated that colonies with queens vaccinated using an experimental *Paenibacillus larvae* bacterin have significantly reduced DWV–B loads compared to colonies with unvaccinated queens, and that this reduction is independent of *Varroa* load.

Deformed Wing Virus (DWV) is one of the most widespread and destructive diseases affecting honey bee colonies (1–3), with the APHIS honey bee survey finding DWV present in over 85% of US hives since 2013 (4). DWV is split into 3 'Master' variants: the common DWV–A (5) and DWV–B (6), and the less common, DWV–C (7). While DWV–A has historically been most prevalent, DWV–B is rapidly overtaking it worldwide (8). Both DWV–A and DWV–B have been directly linked to overwintering failure (3,9), and are easily identified in the hive due to their most striking clinical symptom – undeveloped and non-functional wings on newly emerged adult bees. Bees displaying clinical DWV symptoms are typically never able to forage and die in less than a week after emerging (2).

All DWV variants propagate principally within developing honey bee pupae and are highly associated with the obligate honey bee ectoparasitic mite *Varroa destructor* (10). *V. destructor* mites act as vectors to facilitate indirect horizontal transmission of DWV between pupae (11), and increase the severity of infection by weakening their hosts immune system (12). *V. destructor* are themselves highly associated with colony loss (13) and are often

listed as the factor of greatest concern among beekeepers (14). DWV remains an issue even in the absence of *V. destructor*, as it can be vertically transmitted in from queen to brood, or by direct horizontal transmission to larvae through food (2).

Despite its prevalence and virulence, there is no specific treatment or preventative for DWV (15). Currently, the primary method of reducing DWV levels in a colony is to reduce *V. destructor* levels (16). This is generally done using acaricides and insecticides (17) which stresses the bees (13) and contaminates honey (18). Additionally, reduction of mites does not eliminate DWV, as mites have no impact on vertical or direct horizontal transmission. It has been hypothesized that Trans-Generational Immune Priming (TGIP) using live or inactivated DWV virus could bolster honey bee immunity against DWV, though results to date are mixed (19,20).

**DWV–B Levels Are Significantly Reduced in Honey Bee Colonies With Queens Vaccinated Using Experimental *Paenibacillus larvae* Bacterin, Independent of *V. destructor* Levels**

We placed queens vaccinated with an

experimental *Paenibacillus larvae* bacterin into 200 colonies across 8 sites, alongside an equal number of unvaccinated control colonies, in a large commercial beekeeping operation in Georgia, USA. Vaccinate and control colonies were managed identically for an entire season. All colonies were treated for mites once per month with Amitraz and supplemented with sugar water as necessary according to local nectar availability. Mite counts were taken one week prior to vaccination (N=35 control, N=38 vaccinated) and 6 months after vaccination (N=44 control, N=44 vaccinated) by alcohol washing 300 bees taken from above brood comb. Additionally, pooled samples of adult bees from 10 hives per treatment at each of the 8 yards were taken one week before vaccination (N=8 control, 8 vaccinated), and four months after vaccination (N=8 control, 8 vaccinated). Bee samples were sent to the National Agricultural Genotyping center (21) to have DWV-B viral loads quantified by qPCR.

We found no difference in mite counts between treatments either before (Welch two sample t-test,  $T=0.38$ ,  $df=61$   $P=0.71$ ) or after vaccination (Welch two sample t-test,  $T=-1.30$ ,  $df=48$   $P=0.20$ ) (Figure 1). Mite counts were under 2% (6/300 bees) in >85% of sampled colonies in each treatment at both timepoints.

We found that DWV-B levels were identical between groups prior to vaccination (Wilcoxon rank sum test,  $W=31$ ,  $P=0.96$ ), and were significantly reduced in vaccinated hives compared to control hives 4 months after vaccination (Wilcoxon rank sum test,  $W=54.5$ ,  $P=0.021$ ). DWV-B quantities were reduced in all 8 yards, with an average reduction of 83% (Figure 2). To our knowledge, these data represent the first use of TGIP with a bacterial vaccine to provide protection against a virus in an invertebrate, and the first field example of direct control of DWV over an entire season, independent of mite control.\*

This study demonstrates vaccinating queen

honey bees with *P. larvae* bacterin provides significant cross-protection benefits against DWV-B to honey bee colonies in a commercially-relevant field setting, and that this protection occurs independent of *V. destructor* levels.

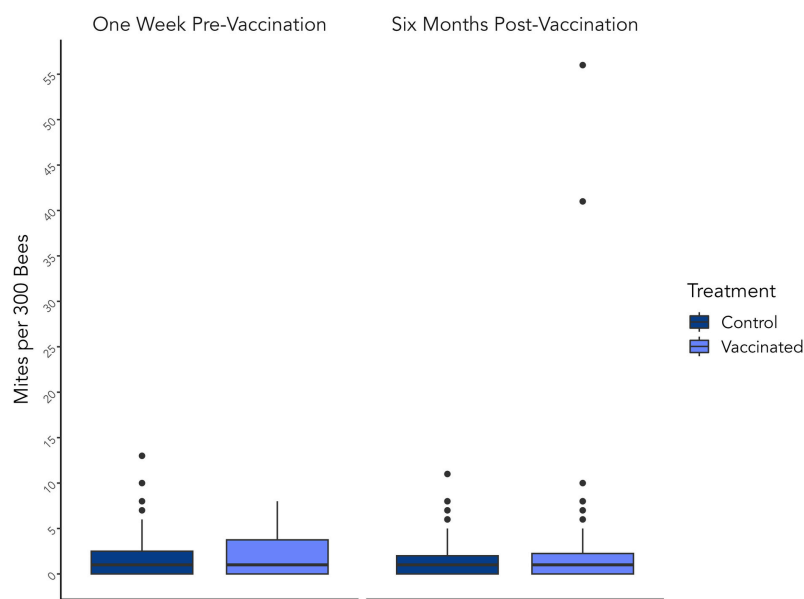
(Figures continue page 3)

*\*The current data do not constitute an efficacy claim for DWV-B for Dalan's existing *P. larvae* product. Further studies and regulatory reviews will be necessary to establish any official claims.*

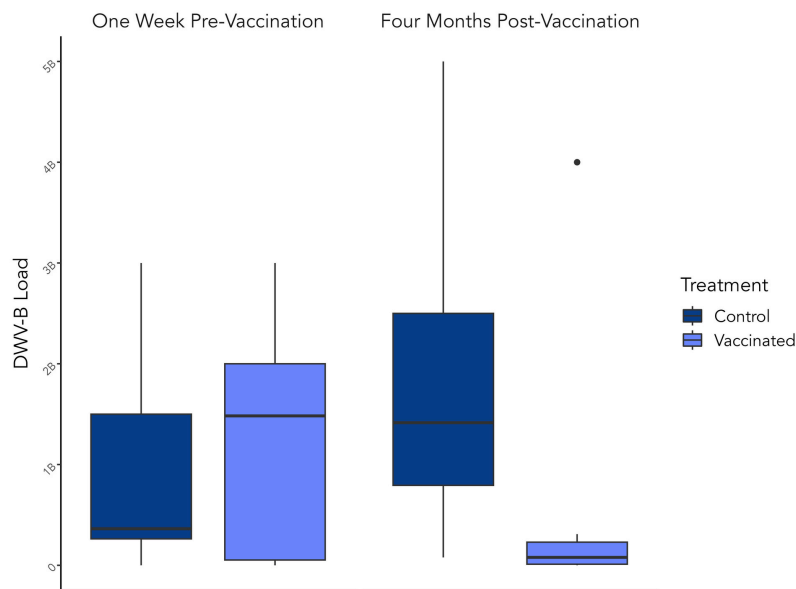
### **About Dalan Animal Health, Inc.**

*Dalan Animal Health is dedicated to bringing the world transformative animal health solutions to support a more sustainable future. This platform vaccine technology uses transgenerational immune priming, allowing the maternal animal to pass immune modulators (e.g., antigens, anti-microbial molecules) to the next generation larvae before they hatch. Dalan plans to develop vaccines for other honeybee diseases and underserved industries, such as shrimp, mealworms, and insects used in agriculture. The company is headquartered in Athens, Georgia, at the University of Georgia's Innovation Hub.*

Figures:



**Figure 1:** Mite count per 300 bees in unvaccinated (navy blue) and vaccinated (light blue) colonies. Bees were collected from above the brood comb and washed for mites one week prior to vaccination (N=35 control, 38 vaccinated colonies), and 6 months post-vaccination (N=44 control, N=44 vaccinated colonies). There was no difference in mite counts between treatments prior to (Welch two sample t-test,  $T=0.38$ ,  $df=61$   $P=0.71$ ), or after vaccination (Welch two sample t-test,  $T=-1.30$ ,  $df=48$   $P=0.20$ ).



**Figure 2:** Box and whisker plot showing DWV-B viral load in pooled bee samples from colonies with either unvaccinated (navy blue) or vaccinated (light blue) queens both before and four months after vaccination. For each treatment, samples were pooled equally from 10 hives in 8 yards (N=8 samples/treatment). Prior to vaccination there was no difference in DWV-B viral load (Wilcoxon rank sum test,  $W=31$ ,  $P=0.96$ ). Four months post vaccination viral load was significantly decreased in vaccinated colonies (Wilcoxon rank sum test,  $W=54.5$ ,  $P=0.021$ ).

## LITERATURE CITED

1. Chen, Y. P. & Siede, R. Honey Bee Viruses. in *Advances in Virus Research* vol. 70 33–80 (Academic Press, 2007).
2. de Miranda, J. R. & Genersch, E. Deformed wing virus. *Journal of Invertebrate Pathology* 103, S48–S61 (2010).
3. Kevill, J. L. et al. DWV–A Lethal to Honey Bees (*Apis mellifera*): A Colony Level Survey of DWV Variants (A, B, and C) in England, Wales, and 32 States across the US. *Viruses* 11, 426 (2019).
4. Bee Informed Partnership – APHIS Survey State Reports. [https://research.beeinformed.org/state\\_reports/](https://research.beeinformed.org/state_reports/).
5. Lanzi, G. et al. Molecular and Biological Characterization of Deformed Wing Virus of Honeybees (*Apis mellifera* L.). *J Virol* 80, 4998–5009 (2006).
6. Ongus, J. R. et al. Complete sequence of a picorna-like virus of the genus Iflavirus replicating in the mite Varroa destructor. *J Gen Virol* 85, 3747–3755 (2004).
7. Mordecai, G. J., Wilfert, L., Martin, S. J., Jones, I. M. & Schroeder, D. C. Diversity in a honey bee pathogen: first report of a third master variant of the Deformed Wing Virus quasispecies. *ISME J* 10, 1264–1273 (2016).
8. Paxton, R. J. et al. Epidemiology of a major honey bee pathogen, deformed wing virus: potential worldwide replacement of genotype A by genotype B. *International Journal for Parasitology: Parasites and Wildlife* 18, 157–171 (2022).
9. Natsopoulou, M. E. et al. The virulent, emerging genotype B of Deformed wing virus is closely linked to overwinter honeybee worker loss. *Sci Rep* 7, 5242 (2017).
10. Martin, S. J., Ball, B. V. & Carreck, N. L. Prevalence and persistence of deformed wing virus (DWV) in untreated or acaricide-treated Varroa destructor infested honey bee (*Apis mellifera*) colonies. *Journal of Apicultural Research* 49, 72–79 (2010).
11. Posada-Florez, F. et al. Deformed wing virus type A, a major honey bee pathogen, is vectored by the mite Varroa destructor in a non-propagative manner. *Sci Rep* 9, 12445 (2019).
12. Nazzi, F. et al. Synergistic parasite–pathogen interactions mediated by host immunity can drive the collapse of honeybee colonies. *PLoS Pathog* 8, e1002735 (2012).
13. Barroso-Arévalo, S. et al. High Load of Deformed Wing Virus and Varroa destructor Infestation Are Related to Weakness of Honey Bee Colonies in Southern Spain. *Frontiers in Microbiology* 10, (2019).
14. Engebretson, J. M. et al. Perceptions of honey bee management information sources among backyard and sideliner beekeepers in the United States. *Journal of Rural Studies* 96, 190–197 (2022).
15. Smeele, Z. E., Baty, J. W. & Lester, P. J. Effects of Deformed Wing Virus–Targeting dsRNA on Viral Loads in Bees Parasitised and Non-Parasitised by Varroa destructor. *Viruses* 15, 2259 (2023).
16. Woodford, L. et al. Quantitative and Qualitative Changes in the Deformed Wing Virus Population in Honey Bees Associated with the Introduction or Removal of Varroa destructor. *Viruses* 14, 1597 (2022).
17. Warner, S. et al. A scoping review on the effects of Varroa mite (*Varroa destructor*) on global honey bee decline. *Science of The Total Environment* 906, 167492 (2024).
18. Pohorecka, K. et al. Amitraz Marker Residues in Honey from Honeybee Colonies Treated with Apiwarol. *J Vet Res* 62, 297–301 (2018).
19. Lang, S., Simone-Finstrom, M. & Healy, K. Context-Dependent Viral Transgenerational Immune Priming in Honey Bees (Hymenoptera: Apidae). *J Insect Sci* 22, 19 (2022).
20. Leponiemi, M., Amdam, G. V. & Freitak, D. Exposure to Inactivated Deformed Wing Virus Leads to Trans-Generational Costs but Not Immune Priming in Honeybees (*Apis mellifera*). *Frontiers in Ecology and Evolution* 9, (2021).
21. Oneil, M. NAGC Launches “BeeCare” Testing for Honey Bee Diseases | NAGC. National Agricultural Genotyping Center <https://genotypingcenter.com/nagc-launches-beecare-testing-for-honey-bee-diseases/> (2017).